

# Iron toxicity, lipid peroxidation and ferroptosis after intracerebral haemorrhage

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## ABSTRACT

Intracerebral haemorrhage (ICH) is a devastating type of stroke with high mortality and morbidity. However, we have few options for ICH therapy and limited knowledge about post-ICH neuronal death and related mechanisms. In the aftermath of ICH, iron overload within the perihematoma region can induce lethal reactive oxygen species (ROS) production and lipid peroxidation, which contribute to secondary brain injury. Indeed, iron chelation therapy has shown efficacy in preclinical ICH studies. Recently, an iron-dependent form of non-apoptotic cell death known as ferroptosis was identified. It is characterised by an accumulation of iron-induced lipid ROS, which leads to intracellular oxidative stress. The ROS cause damage to nucleic acids, proteins and lipid membranes, and eventually cell death. Recently, we and others discovered that ferroptosis does occur after haemorrhagic stroke in vitro and in vivo and contributes to neuronal death. Inhibition of ferroptosis is beneficial in several in vivo and in vitro ICH conditions. This minireview summarises current research on iron toxicity, lipid peroxidation and ferroptosis in the pathomechanisms of ICH, the underlying molecular mechanisms of ferroptosis and the potential for combined therapeutic strategies. Understanding the role of ferroptosis after ICH will provide a vital foundation for cell death-based ICH treatment and prevention.

Intracerebral haemorrhage (ICH) accounts for 10%–15% of all strokes. It is a devastating event that causes high mortality and morbidity, but it currently lacks effective therapies.<sup>1</sup> ICH is estimated to affect over 1 million people worldwide each year, and the number is expected to rise substantially as the population ages. Most patients with ICH die or become severely disabled, and the mainstay for current therapy is supportive care. Consequently, only 20% of survivors have functional independence at 6 months.<sup>2,3</sup> Primary brain damage develops in the first few hours after ICH as the formation and expansion of haematoma or oedema induce mass effects and elevations in intracranial pressure that lead to herniation and death. In the aftermath of ICH, iron accumulation within the perihematoma region contributes to the secondary brain damage.<sup>4–12</sup>

ICH occurs when a weakened vessel ruptures and bleeds into the surrounding

brain.<sup>6,13</sup> The blood accumulates and compresses the surrounding brain tissue, causing tissue damage, inflammation and neuronal death.<sup>14–16</sup> Preclinical and clinical studies suggest that toxins released from an intracerebral haematoma may contribute to brain damage after ICH.<sup>5–8,12–17</sup> Haemoglobin (Hb)/haem is one putative neurotoxin. Hb, the most abundant protein in blood, is released from lysed red blood cells after ICH. It can be engulfed by microglia and infiltrating macrophages in the perihematoma zone and metabolised into ferrous/ferric iron, which induces the formation of lethal reactive oxygen species (ROS) and lipid peroxidation.<sup>8–12</sup> Subsequently, excess ferrous iron is transported out of microglia, accumulates in neurons via the transferrin (Tf)–Tf receptor system and reacts with hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) via the Fenton reaction to form highly toxic hydroxyl radicals (•OH) (figure 1). These hydroxyl radicals attack DNA, proteins and lipid membranes, thereby disrupting cellular function.<sup>18</sup> In our previous studies, we showed that iron toxicity contributes to ICH-induced early brain injury and that reducing iron accumulation with iron chelators (deferoxamine, 2,2'-dipyridyl, or VK-28) reduces grey and white matter injury and improves functional outcomes.<sup>8,9,11</sup> Additionally, we were the first group to demonstrate that an iron-dependent form of non-apoptotic cell death known as ferroptosis occurs in a mouse model of ICH.<sup>10</sup> Therefore, patients with ICH might benefit from a reduction in Hb/iron-induced toxicity or direct neuronal rescue.<sup>6,12</sup>

Researchers have identified various forms of cell death after ICH, including apoptosis<sup>19–24</sup> and necrosis<sup>20,25</sup> in humans and experimental animals, and autophagic cell death<sup>26,27</sup> in animal models. Whereas mitochondria are usually swollen in each of these forms of cell death, ferroptotic cells exhibit mitochondrial shrinkage. In their 2016 review, Xie *et al.*<sup>28</sup> provide direct comparisons of the morphological, biochemical and immune features of

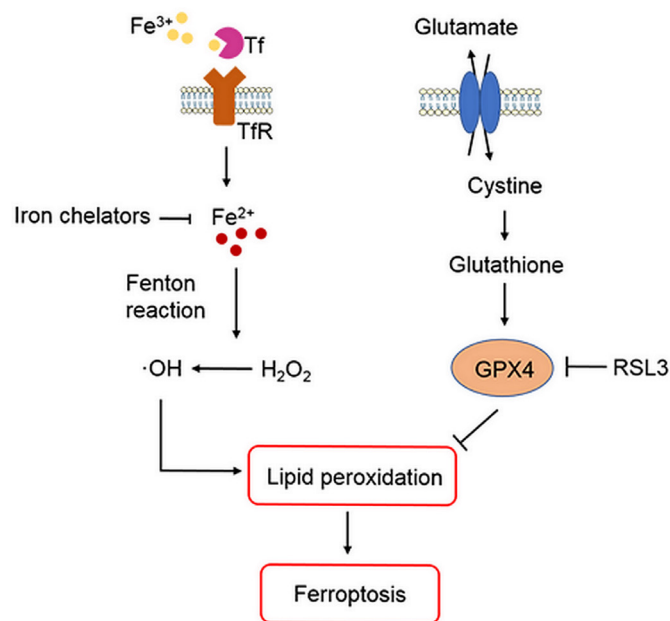


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**Figure 1** Role of iron, glutathione and lipid peroxidation in ferroptosis. Arrows indicate promotion; blunt-ended lines indicate inhibition. GPX4, glutathione peroxidase 4;  $\text{H}_2\text{O}_2$ , hydrogen peroxide;  $\bullet\text{OH}$ , hydroxyl radicals; RSL3, Ras-selective lethal 3; Tf, transferrin; TfR, transferrin receptor.

ferroptosis, apoptosis, necrosis and autophagy. Inhibition of apoptosis, necrosis and autophagy improve outcomes in ICH animal models.<sup>22 25 27</sup> However, caspase inhibitors have failed to inhibit neuronal death induced by Hb,<sup>29</sup> and multiple forms of cell death may contribute collectively to neuronal death after ICH. Indeed, Zhu *et al.*<sup>25</sup> showed that necrosis occurs earlier than apoptosis but that both contribute to cell death after ICH. Others have reported that both apoptosis and necrosis are present in brain sections from patients with ICH.<sup>20 24</sup> These results prompted us to investigate whether ferroptotic cell death occurs after ICH. In a recent study, ferroptosis was identified in cancer cells and in organotypic hippocampal slice cultures (OHSCs) exposed to glutamate.<sup>30</sup> Ferroptosis is distinguishable from other forms of regulated cell death because it does not require caspases (mediators of apoptosis and pyroptosis), ATP depletion or mitochondrial ROS generation (mediators of necroptosis), Bax/Bak (essential mediators of mitochondrial outer membrane permeabilization), or elevations in intracellular  $\text{Ca}^{2+}$ .<sup>30</sup> Ferroptosis is triggered by glutathione biosynthesis or glutathione peroxidase 4 (GPX4) activity inhibition<sup>28 31–33</sup> (figure 1) and is associated morphologically with shrunken and electron-dense mitochondria.<sup>30</sup> Three mechanisms in cancer cells have been reported to lead to ferroptosis: (1) depletion of glutathione and subsequent inactivation of GPX4, (2) Tf import or iron overload and (3) glutamine metabolism.<sup>30 31 34</sup> Therefore, intracellular iron overload is clearly a major mechanism of Hb-induced and ferrous iron-induced neuronal ferroptosis after ICH, and decreasing iron accumulation in microglia might attenuate neuronal death. In a middle cerebral artery

occlusion model of ischaemic stroke, ferroptosis contributes to neuronal death through tau-iron interaction.<sup>35</sup> In addition to haemorrhagic and ischaemic stroke, ferroptosis has been shown to have significant implications in several other neurological diseases, such as Alzheimer's disease, Parkinson's disease and brain tumours.<sup>36</sup>

We have previously shown that ferroptosis contributes to neuronal death in the mouse collagenase-induced ICH model.<sup>10</sup> Using transmission electron microscopy, we observed shrunken mitochondria in the brains of ICH mice, providing powerful evidence of ferroptosis. In OHSCs, we found that administration of ferrostatin-1 (Fer-1), a specific inhibitor of ferroptosis, prevented neuronal death and reduced iron deposition induced by Hb. In vivo, mice treated with Fer-1 after ICH showed marked brain protection and improved neurological function. We also found that Fer-1 reduced lipid ROS production and attenuated the increased expression level of prostaglandin-endoperoxide synthase 2 (PTGS2) and its gene product cyclooxygenase-2 (COX-2) in OHSCs and in vivo. Notably, COX-2 is highly expressed in neurons after ICH, and inhibition of COX-2 reduces ICH-induced secondary brain injury.<sup>37 38</sup> Therefore, COX-2 might be a biomarker of ferroptosis. Based on these findings, we investigated whether a combination of cell death inhibitors could improve neuronal rescue. We found that Fer-1 in combination with other inhibitors that target different forms of cell death prevented Hb-induced cell death in OHSCs and human-induced pluripotent stem cell-derived neurons better than any inhibitor alone.<sup>10</sup> Our most recent study showed that ferroptosis coexists with necrosis and autophagy in the mouse ICH brain.<sup>39</sup> Therefore, it should be further determined whether a combination of inhibitors can improve ICH outcomes in animals better than one drug, and whether this strategy can be effective in clinical trials. In another new study, Zhang *et al.*<sup>40</sup> showed that GPX4 expression was markedly reduced during the acute phase of ICH and that increasing GPX4 level was able to rescue neurons from ferroptotic death and improve outcomes in rats. These novel findings fill an important gap in knowledge regarding cell death after ICH and provide a vital foundation for cell death-based ICH treatment in the future.

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## REFERENCES

- Donnan GA, Hankey GJ, Davis SM. Intracerebral haemorrhage: a need for more data and new research directions. *Lancet Neurol* 2010;9:133–4.
- Hemphill JC, Greenberg SM, Anderson CS, et al. Guidelines for the management of spontaneous intracerebral hemorrhage: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2015;46:2032–60.
- Thabet AM, Kottapally M, Hemphill JC. Management of intracerebral hemorrhage. *Handb Clin Neurol* 2017;140:177–94.
- Wu J, Hua Y, Keep RF, et al. Iron and iron-handling proteins in the brain after intracerebral hemorrhage. *Stroke* 2003;34:2964–9.
- Xi G, Keep RF, Hoff JT. Mechanisms of brain injury after intracerebral haemorrhage. *Lancet Neurol* 2006;5:53–63.
- Wang J. Preclinical and clinical research on inflammation after intracerebral hemorrhage. *Prog Neurobiol* 2010;92:463–77.
- Lou M, Lieb K, Selim M. The relationship between hematoma iron content and perihematoma edema: an MRI study. *Cerebrovasc Dis* 2009;27:266–71.
- Wu H, Wu T, Xu X, et al. Iron toxicity in mice with collagenase-induced intracerebral hemorrhage. *J Cereb Blood Flow Metab* 2011;31:1243–50.
- Wu H, Wu T, Li M, et al. Efficacy of the lipid-soluble iron chelator 2,2'-dipyridyl against hemorrhagic brain injury. *Neurobiol Dis* 2012;45:388–94.
- Li Q, Han X, Lan X, et al. Inhibition of neuronal ferroptosis protects hemorrhagic brain. *JCI Insight* 2017;2:e90777.
- Li Q, Wan J, Lan X, et al. Neuroprotection of brain-permeable iron chelator VK-28 against intracerebral hemorrhage in mice. *J Cereb Blood Flow Metab* 2017;37:3110–23.
- Xiong XY, Wang J, Qian ZM, et al. Iron and intracerebral hemorrhage: from mechanism to translation. *Transl Stroke Res* 2014;5:429–41.
- Wang J, Tsirka SE. Contribution of extracellular proteolysis and microglia to intracerebral hemorrhage. *Neurocrit Care* 2005;3:077–85.
- Wang J, Doré S. Inflammation after intracerebral hemorrhage. *J Cereb Blood Flow Metab* 2007;27:894–908.
- Wasserman JK, Schlichter LC. Neuron death and inflammation in a rat model of intracerebral hemorrhage: effects of delayed minocycline treatment. *Brain Res* 2007;1136:208–18.
- Wu H, Zhang Z, Hu X, et al. Dynamic changes of inflammatory markers in brain after hemorrhagic stroke in humans: a postmortem study. *Brain Res* 2010;1342:111–7.
- Xi G, Strahle J, Hua Y, et al. Progress in translational research on intracerebral hemorrhage: is there an end in sight? *Prog Neurobiol* 2014;115:45–63.
- Salvador GA. Iron in neuronal function and dysfunction. *Biofactors* 2010;36:103–10.
- Matsushita K, Meng W, Wang X, et al. Evidence for apoptosis after intercerebral hemorrhage in rat striatum. *J Cereb Blood Flow Metab* 2000;20:396–404.
- Qureshi AI, Suri MF, Ostrow PT, et al. Apoptosis as a form of cell death in intracerebral hemorrhage. *Neurosurgery* 2003;52:1041–7.
- Gong C, Boulis N, Qian J, et al. Intracerebral hemorrhage-induced neuronal death. *Neurosurgery* 2001;48:875–82.
- Grossetete M, Rosenberg GA. Matrix metalloproteinase inhibition facilitates cell death in intracerebral hemorrhage in mouse. *J Cereb Blood Flow Metab* 2008;28:752–63.
- Serrano L, Martínez-Redondo P, Marazuela-Duque A, et al. The tumor suppressor Sirt2 regulates cell cycle progression and genome stability by modulating the mitotic deposition of H4K20 methylation. *Genes Dev* 2013;27:639–53.
- Zhang XQ, Zhang ZM, Yin XL, et al. Exploring the optimal operation time for patients with hypertensive intracerebral hemorrhage: tracking the expression and progress of cell apoptosis of prehematoma brain tissues. *Chin Med J* 2010;123:1246–50.
- Zhu X, Tao L, Tejima-Mandeville E, et al. Plasmalemma permeability and necrotic cell death phenotypes after intracerebral hemorrhage in mice. *Stroke* 2012;43:524–31.
- He Y, Wan S, Hua Y, et al. Autophagy after experimental intracerebral hemorrhage. *J Cereb Blood Flow Metab* 2008;28:897–905.
- Chen CW, Chen TY, Tsai KL, et al. Inhibition of autophagy as a therapeutic strategy of iron-induced brain injury after hemorrhage. *Autophagy* 2012;8:1510–20.
- Xie Y, Hou W, Song X, et al. Ferroptosis: process and function. *Cell Death Differ* 2016;23:369–79.
- Wang X, Mori T, Sumii T, et al. Hemoglobin-induced cytotoxicity in rat cerebral cortical neurons: caspase activation and oxidative stress. *Stroke* 2002;33:1882–8.
- Dixon SJ, Lemberg KM, Lamprecht MR, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* 2012;149:1060–72.
- Yang WS, SriRamaratnam R, Welsch ME, et al. Regulation of ferroptotic cancer cell death by GPX4. *Cell* 2014;156:317–31.
- Cao JY, Dixon SJ. Mechanisms of ferroptosis. *Cell Mol Life Sci* 2016;73:2195–209.
- Friedmann Angeli JP, Schneider M, Proneth B, et al. Inactivation of the ferroptosis regulator Gpx4 triggers acute renal failure in mice. *Nat Cell Biol* 2014;16:1180–91.
- Gao M, Monian P, Quadri N, et al. Glutaminolysis and transferrin regulate ferroptosis. *Mol Cell* 2015;59:298–308.
- Tuo QZ, Lei P, Jackman KA, et al. Tau-mediated iron export prevents ferroptotic damage after ischemic stroke. *Mol Psychiatry* 2017;22:1520–30.
- Weiland A, Wang Y, Wu W, et al. Ferroptosis and its role in diverse brain diseases. *Mol Neurobiol* 2018.
- Chu K, Jeong SW, Jung KH, et al. Celecoxib induces functional recovery after intracerebral hemorrhage with reduction of brain edema and perihematoma cell death. *J Cereb Blood Flow Metab* 2004;24:926–33.
- Wu T, Wu H, Wang J, et al. Expression and cellular localization of cyclooxygenases and prostaglandin E synthases in the hemorrhagic brain. *J Neuroinflammation* 2011;8:22.
- Li Q, Weiland A, Chen X, et al. Ultrastructural characteristics of neuronal death and white matter injury in mouse brain tissues after intracerebral hemorrhage: coexistence of ferroptosis, autophagy, and necrosis. *Front Neurol* 2018;9:581.
- Zhang Z, Wu Y, Yuan S, et al. Glutathione peroxidase 4 participates in secondary brain injury through mediating ferroptosis in a rat model of intracerebral hemorrhage. *Brain Res* 2018;1701:112–25.